

ourse: Optics, forces & development

In vivo 3D-microscopy for the analysis of cell behaviour in developing embryos

Santiago - Chile, January 14-29th, 2013

This practical and theoretical course is aimed at graduate students from Latin America interested in the use of optics and microscopic techniques for in vivo 3D visualisation and analysis of cell and tissue dynamics.

Topics:

Development of zebrafish and annual fish Confocal and spinning microscopy Light-sheet microscopy Super-resolution microscopy Photoactivation and laser ablation In vivo electroporation Force estimation in cells & tissues Optical tweezers Particle tracking Image processing and onalysis

Teachers:

Roberto Bernal (U. Santiago, Chi) Sebastián Brauchi (U. Austral, Chi) Miguel Concha (U. Chile, Chil Mauricio Cerda (U. Chile, Chi) Jorg Enderlein (Univ. Gottingen, Ger) Nikta Fakhri (Univ. Gottingen, Ger) Steffen Härtel (U. Chile, Chil Carl-Philipp Heisenberg (IST. Austria) Jorge Jara (U. Chile, Chi) Ulrich Kubitscheck (Univ. Bonn, Ger) Omar Ramírez (U. Chile, Chi) Florian Rehfeldt (Univ. Gottingen, Gerl German Reig (U. Chile, Chi) Felipe Santibañez (U. Chile, Chil Christoph Schmidt (Univ. Gottingen, Ger) . Jan Spille (Univ. Bonn, Ger) Juan Pablo Staforelli (CEFOP, Chi)

Organisers:

. Miguel Concha (U. Chile) . Steffen Härtel (U. Chile)

Travel Fellowships will be available. Indicate your interest in the application.



BIOMEDICAL NEUROSCIENCE INSTITUTE





Informaciones en:

-> <u>www.scian.cl</u>

Notas:

Prácticos (25 %) Seminarios (25 %) Examen Final (50%)

3 grupos de 2/4 para:

- el seminario:

90 min presentación, preguntas y discusión







Optics, Forces & Development 14.01 – 29.01 | 2013

Steffen Härtel

Programa de Anatomía y Biología del Desarollo, BNI, ICBM, Facultad de Medicina, Universidad de Chile, Santiago, Chile







Optics, Forces & Development 14.01 – 29.01 | 2013

Steffen Härtel

- Introduction to microcopy for in vitro and in vivo imaging

- Theory of fluorescence. Diffraction theory & diffraction limited resolution. Point Spread Function and convolution

- Microscopy Acquisition: Nyquist / Sampling Theorem. Signal/noise ratio



















"It is very easy to answer many of these fundamental biological questions. You just look at the thing !

Make microscopes a hundred times more powerful and many problems of biology would be made very much easier."

Richard Feynman (1918-1988)







... just look at the thing ... ¿ Human visual perception ?

Treatise of man (~ 1637)

Passions of the soul (~ 1649)

René Descartes (1596-1650)







glandula pinealis / pineal organ

A combination of ...

- 1| direct signals ...
- 2| signals from other senses ...
- 3| feedback loops ...

... produce a symbolic representation of an object.













Langmuir trough coupled to time lapse microscopy









Fluorescence microscopy: visualization













Over- and Under-sampling

BIOMEDICAL

NSTITUTE

EUROSCIENCE





Richard W Cole¹, Tushare Jinadasa² & Claire M Brown^{2,3}













Spline interpolation















Spline interpolation



Parametric curve:

 $C{=}C(s{,}t)=[x(s{,}t){,}y(s{,}t)]$





















BIOMEDICAL

INSTITUTE

NEUROSCIENCE





Parametrización







Parametrización











Fuerzas



diploes -> dipole moment density -> inter domain energy

5 = 0 0 = 0 $2 \times 10^{-20} 4 \times 10^{-20} 6 \times 10^{-20} 8 \times 10^{-20} 1 \times 10^{-19}$ N = 742 |-> Inter Domain Dipole Energy



Fuerzas

















noi% [Cer] 5 n	nol% [Cer]		20 mol% [Cer] 30 mol% [Cer]			50 mol% [Ce	
200	300	400	500	↓ ₆₀₀	700	800	time [s] -
2 t3 t3'	t3"	*	3×	t3"" t3 ^{xx}	*	t4 t4	t511
←domain crystallization				linear domain growth		contact between	
circula relaxing curvature	1st bord	er undulat co	ion 2nd border undulation nstant border curvature			domain borders	
lattic centre di	e formation / stances incre	ase		hexag centre dis	onal lattice / tances decreas	e	
border distances decrease			decrease	se fast decrease slow			
inter domain energy increases			increases fast increa			ases	
domain mover	ment uncou	oled ro	tationally cou	ipled→	laterally cou	pled→	
lag-time slow beginning an hydrolysis of steady-sta			d 1 st part 2 nd part and ending of steady-state reaction		rate fall and reaction halting		





SM→Cer conversion Mixture of SM / Cer induced by SMase

a: 1.8 mol% Cer	g: 2 mol% Cer
b: 10.6 mol% Cer	h: 10 mol% Cer
c: 20 mol% Cer	i: 20 mol% Cer
d: 25 mol% Cer	j: 25 mol% Cer
e: 32.1 mol% Cer	k: 30 mol% Cer
<u>f: 50 mol% Cer</u>	1: 50 mol% Cer







Laura Fanani



2D · Morpho-topology in lipid monolayers

2010 BBA 2009 Biophysical Journal 2007 Cell Biochemistry and Biophysics 2005 Biophysical Journal 2003 Chemisty and Physics of Lipids 2002 Biophysical Journal

Bruno Maggio



- For enzyme-free SM/Cer monolayers, the LC domains formed cover an area larger than expected. Cer-enriched domains at equilibrium conditions contains about 50% of SM.
- In SMase-domains present a high content of Cer, **higher intradomain repulsion** and, as a consequence, star-like shaped domains, while condensed rounded domains formed in the enzyme-free films.
- The self-organization into highly ordered hexagonal lattice patterns is a consequence of a enhanced interdomain repulsion. **Repulsion is lower in enzyme-free monolayers.**









At T = 20 °C: Solid Gel or S₀ phase is formed by DPPC 16:0 (1,2-Dipalmitoyl-*sn*-Glycero-3-Phosphocholine) + 0.5 mol% $DiIC_{18}$.



At T = 20 °C: Fluid phase is formed by DLPC 12:0 PC (1,2-Dilauroylsn-Glycero-3-Phosphocholine) + 0.5 mol% BODIPY-PC.



Interpolación de contornos







Interpolación de contornos





X_{DPPC}



Interpolación de contornos









Luis Bagatolli (MEMPHYS, DK)



3D · Morpho-topology

2012 European Biophysics Journal 2010 J of Struct Biol 2010 Development 2010 Biological Research **2009 BBA** 2007 Computational Modeling



Miguel Concha



GUV populations can be prepared with excellent reproducibility.

The theoretical lever was approved experimentally in GUVs .

2D-lipid domain surfaces can be determined reliably from 3D image stacks.

Facultad de Medicina, U-Chile

Morpho-genetic mechanisms, form, function in developmental biology

Transgenic *flh*::GFP

Deconvolved Data

morphology

basolateral

Similar for α-tubulin, γ-tubulin, Phalloidin,

$$E = \int_{0}^{1} \frac{1}{2} \left[\alpha \left| \frac{\delta C(s)}{\delta s} \right|^{2} + \beta \left| \frac{\delta^{2} C(s)}{\delta s^{2}} \right|^{2} \right]$$

 $+ E_{ext}[C(s)]ds$

 $+E_n[C(s),C_i(s)]ds$

 $+ E_{f}[C(s), C(s, t+1), C(s, t-1)]ds$

internal energy

contour dependent

(Kass et al 1988)

external energy

image dependent

(GGVF Xu&Price 1988)

distance energy neighbour dependent (us ...) CLG-3D optical flow energy time dependent ++ multi-grid

(Bruhn et al. 2003/2005)

++ 'subjective surfaces'

(Sarti et al 2000)