

Curso Binacional 2023  
Uruguay - Chile

### Microscopía para el Estudio de Biofilms Bacterianos

Instituto de Investigaciones Biológicas Clemente Estable (IIBCE)  
Instituto de Neurociencia Biomédica (BNI), ICBM, F-Med, U-Chile

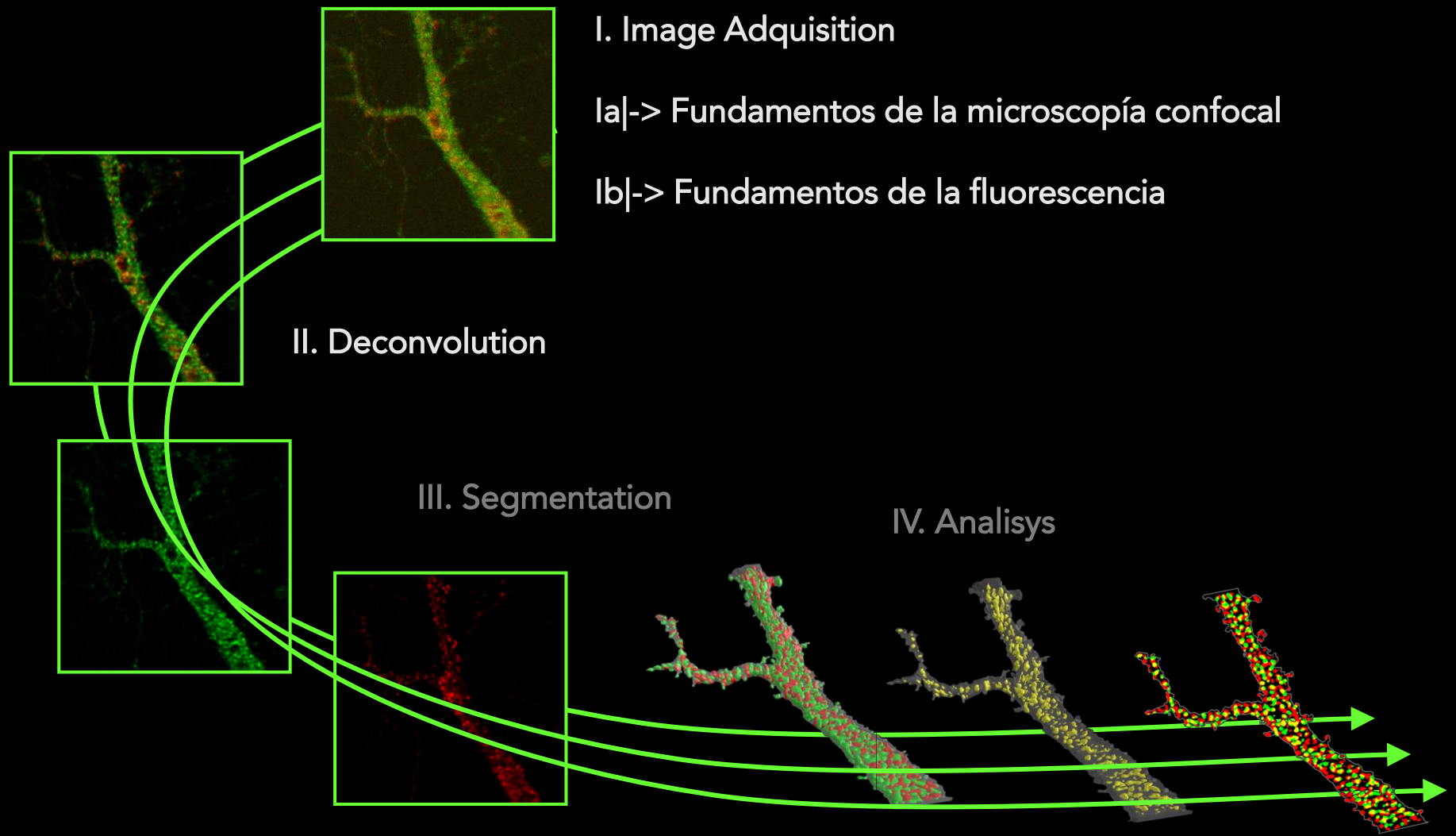
2 - 6 de octubre - Teórico  
9 - 13 de octubre - Práctico Uy,  
10 de octubre - 3 de noviembre - Práctico Cl.



Prof. Dr. Steffen Härtel

[www.scian.cl](http://www.scian.cl) / [www.cimt.cl](http://www.cimt.cl) / [www.cens.cl](http://www.cens.cl) [www.bni.cl](http://www.bni.cl) / [www.rsdue.cl](http://www.rsdue.cl)

Laboratory for Scientific Image Analysis (SCIAN-Lab)  
Centro de Informática Médica y Telemedicina (CIMT)  
Centro Nacional en Sistemas de Información en Salud (CENS)  
Biomedical Neuroscience Institute (BNI)  
Red de Salud Digital de Universidades del Estado (RSDUE)  
Institute of Biomedical Sciences (ICBM)  
Anatomy and Developmental Biology Program  
Escuela de Postgrado  
Facultad de Medicina, Universidad de Chile



- Principles of Fluorescence Spectroscopy, Joseph R. Lakowicz 4.1 Introduction to Fluorescence

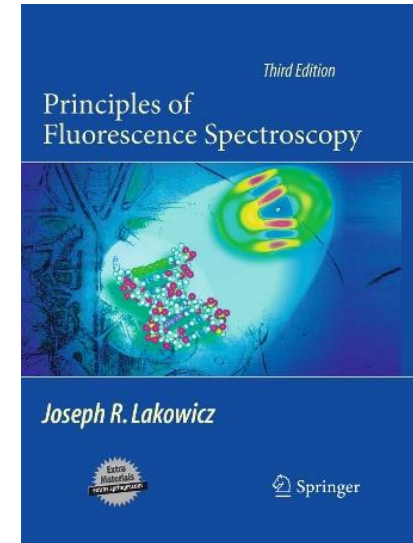
|-> Quenching, Bleaching ...

|-> Polarisation ...

|-> Steady-State and Time-Resolved Fluorescence...

|-> Förster Resonance Energy Transfer ...

- Photomultiplier [www.olympus-lifescience.com/en/microscope-resource/primer/digitalimaging/concepts/photomultipliers](http://www.olympus-lifescience.com/en/microscope-resource/primer/digitalimaging/concepts/photomultipliers)
- Chrome Spectra Viewer [www.chroma.com/spectra-viewer](http://www.chroma.com/spectra-viewer)
- Thermo Fisher Spectra [www.thermofisher.com/order/spectra-viewer](http://www.thermofisher.com/order/spectra-viewer)



## Luminescencia:



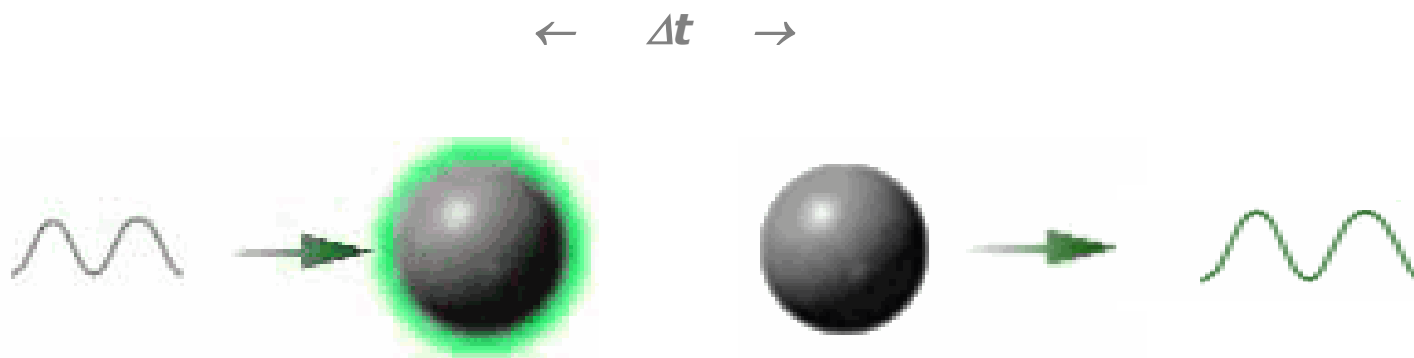
- *Fluorescencia*  $\Delta t \sim 10^{-8}s$
- *Fosforescencia*  $\Delta t \sim 10^{-3}-10^0s$

## Interacciones ...

- intra- e inter moleculares ...

## producen cambios ...

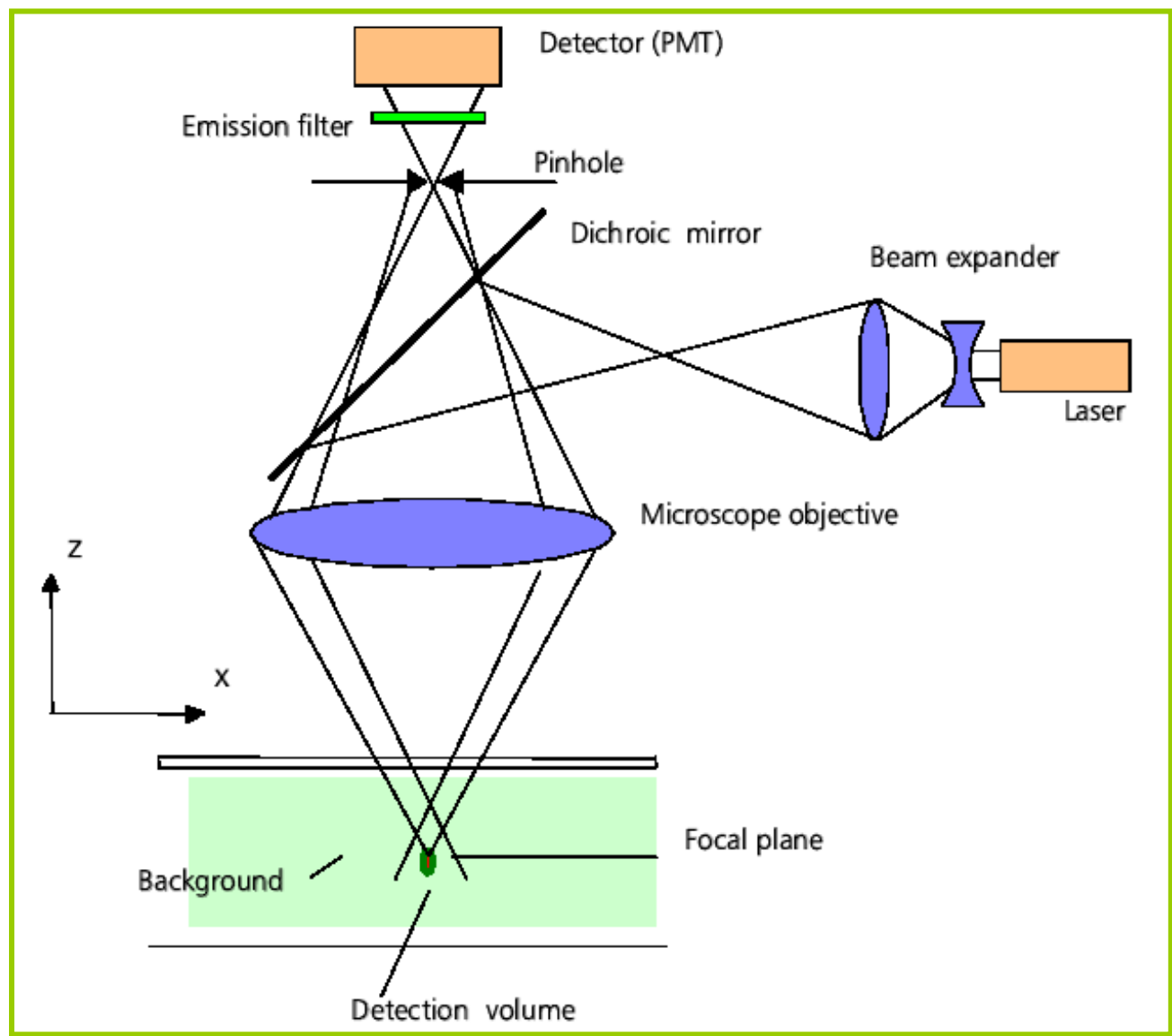
- espectrales
- tiempos de vida
- polarización
- intensidad ...

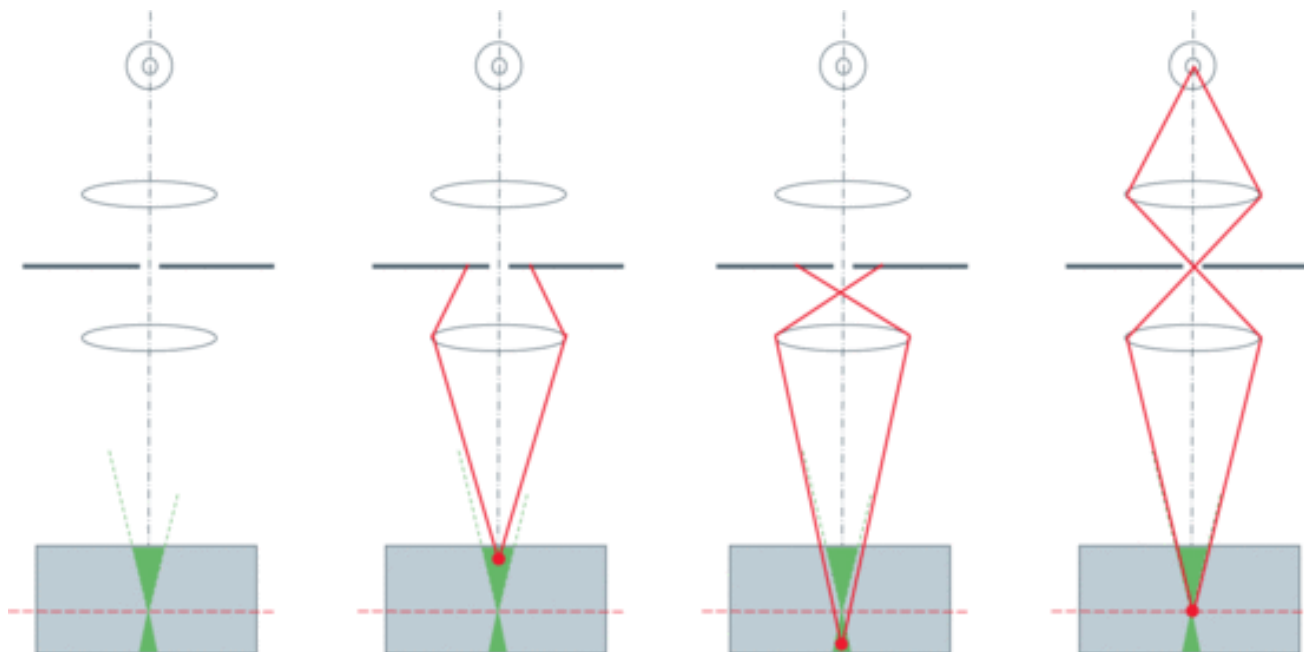


*Absorción / Excitación*

*Emisión*

# | -> Diffraction limited Microscopy

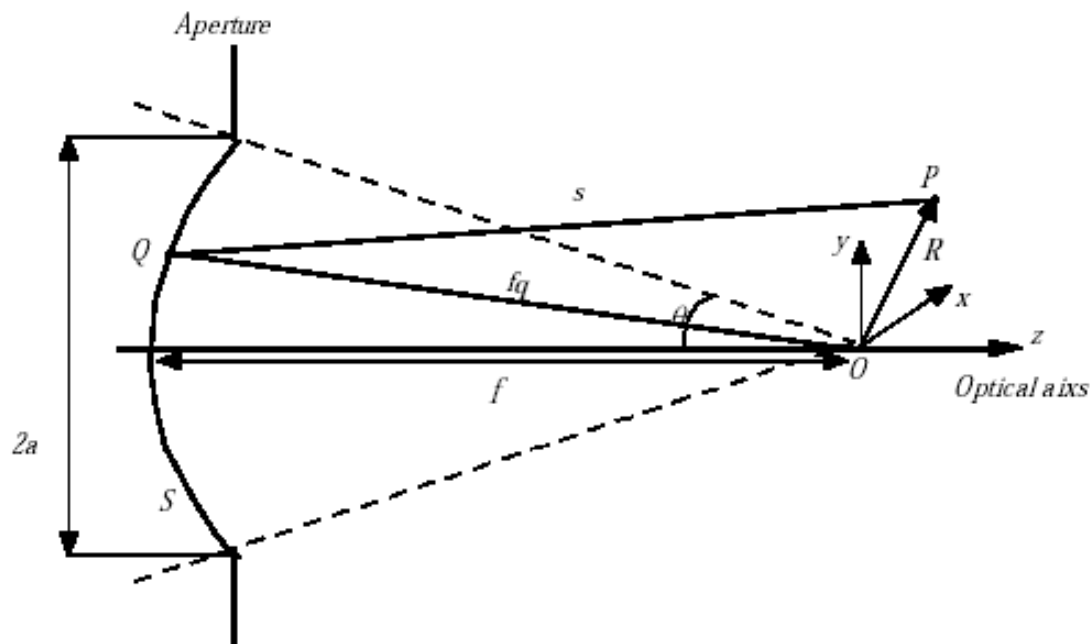




Rebanada óptica en  $\mu\text{m}$ , modificable según Airy units del pinhole

## From Geometric Optics to Diffraction Theory:

**Diffraction:** The deviation of an electromagnetic wavefront from the path predicted by geometric optics when the wavefront interacts with a physical object such as an opening or an edge.



**Figure 2.1** Diffraction of a converging spherical wave at a circular aperture

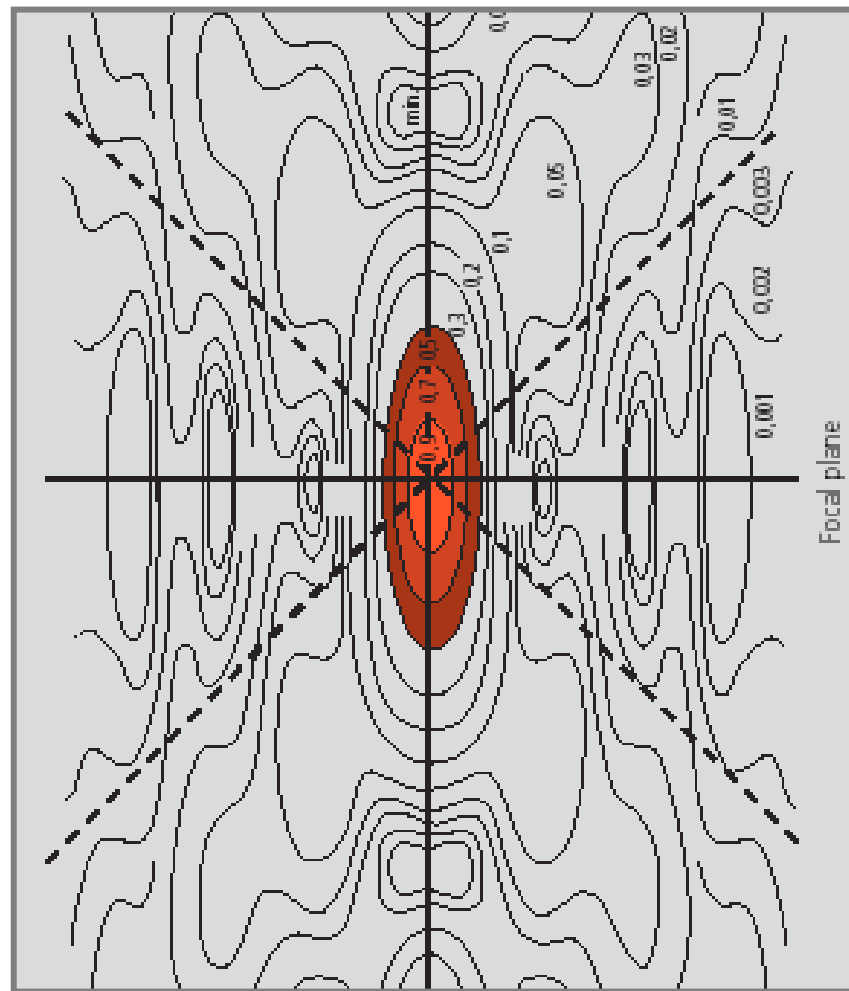
## Óptica no-geométrica / Teoría de difracción

$$\text{PSF} = |U|^2 = f(J_0)$$

$U$ , Integral de Difracción de Kirhoff

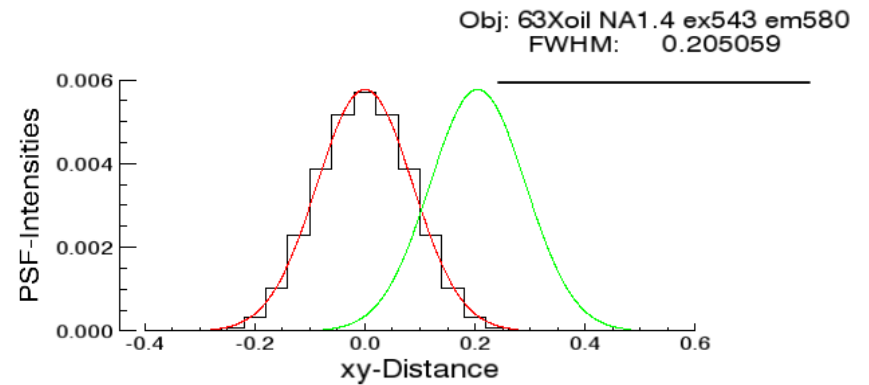
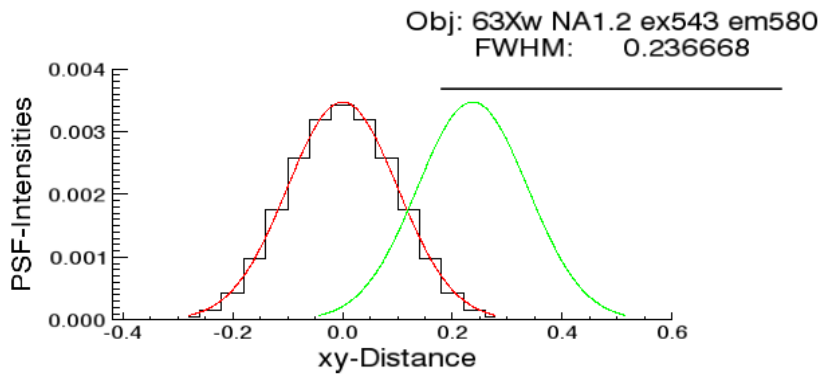
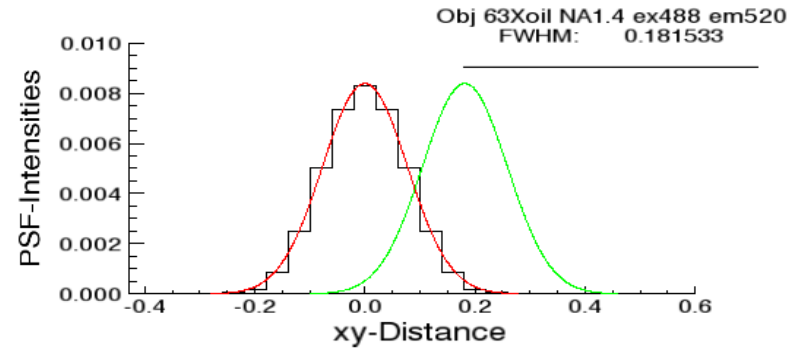
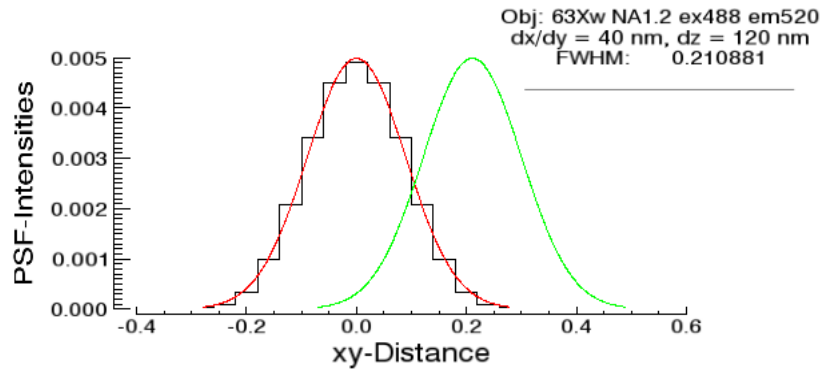
$J_0$ , Serie de funciones de Bessel

*(Born & Wolf, Principles of Optics, 6th edition 1988,  
Pergamon Press)*

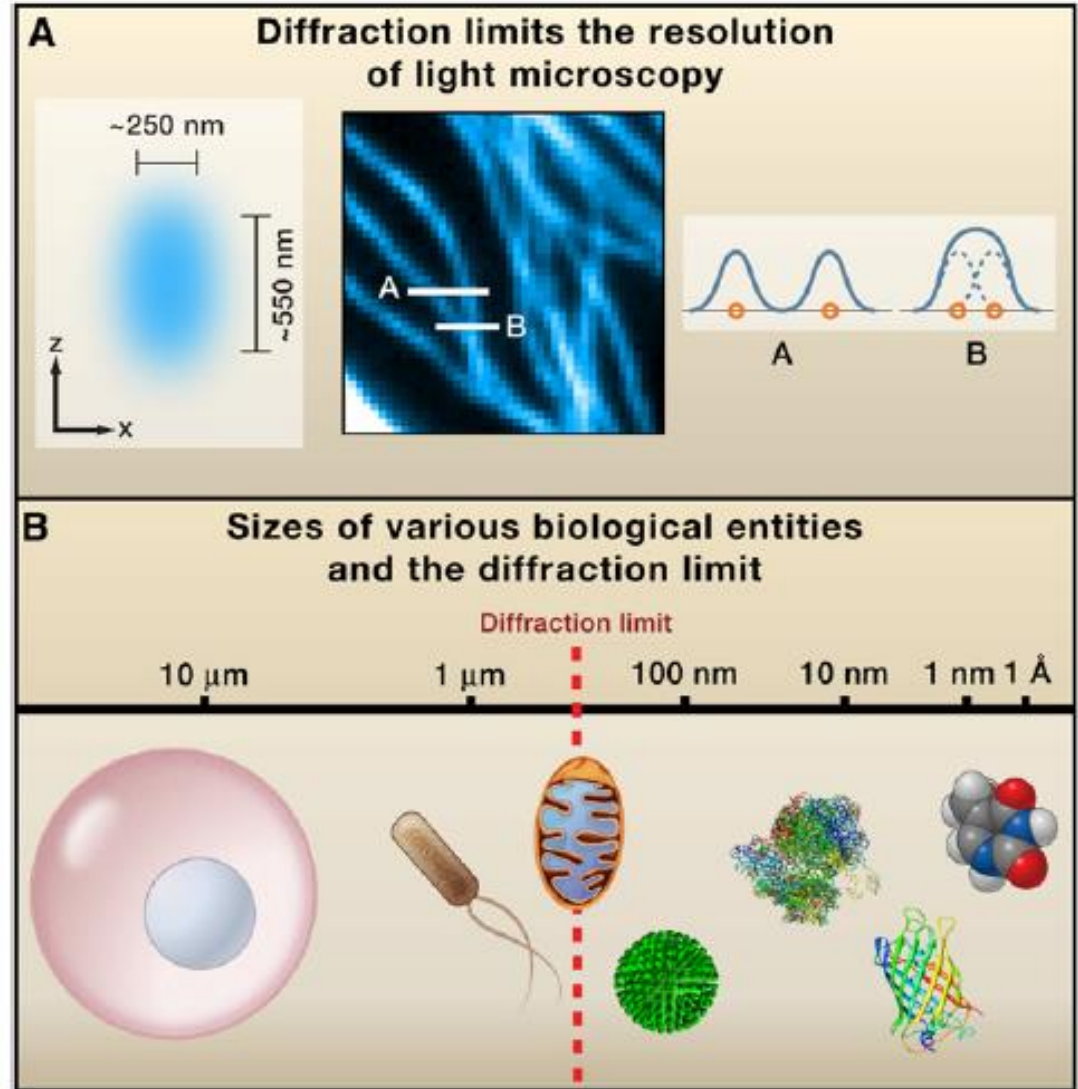
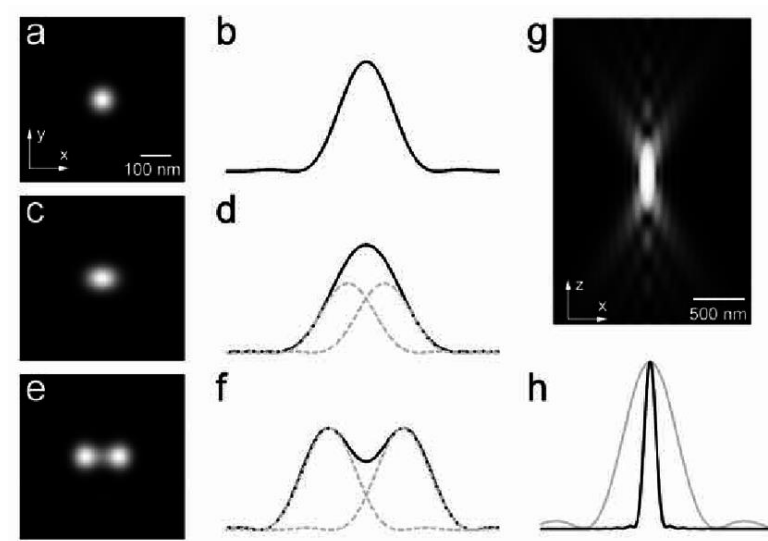




# | -> Diffraction limited Microscopy



# | -> Diffraction limited Microscopy



Bertocchi, Cristina & Goh, Wah & Zhang, Zhen & Kanchanawong, Pakorn. (2013). Nanoscale Imaging by Superresolution Fluorescence Microscopy and Its Emerging Applications in Biomedical Research. *Critical reviews in biomedical engineering*. 41. 281-308. 10.1615/CritRevBiomedEng.2014010448.

# | -> Beyond diffraction

M Goeppert-Mayer  
1906-1972

M Gustafson  
1960-2011

S Hell  
MPI Göttingen  
BIOQUANT Hdg

E Betzig  
Janelia Farm



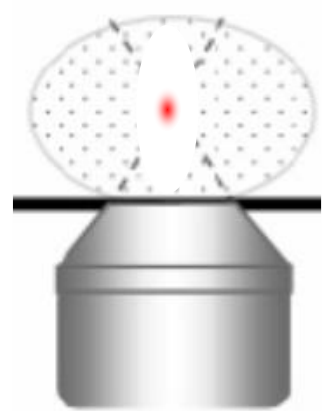
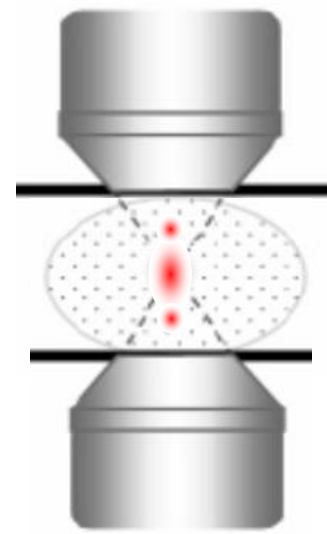
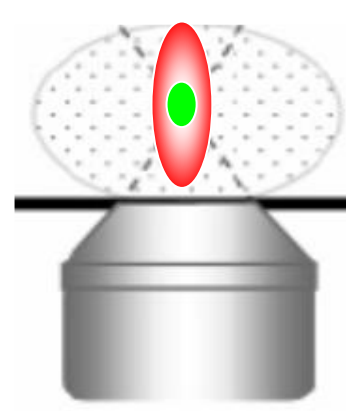
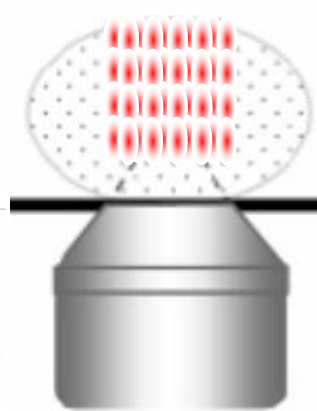
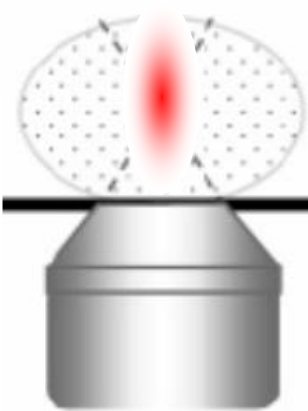
FWHM(xy)  $\sim \lambda/2$

$\sim \lambda/4$

$\sim \lambda/\infty$

$\sim \lambda/4$

$\sim \lambda/100$



2-photon

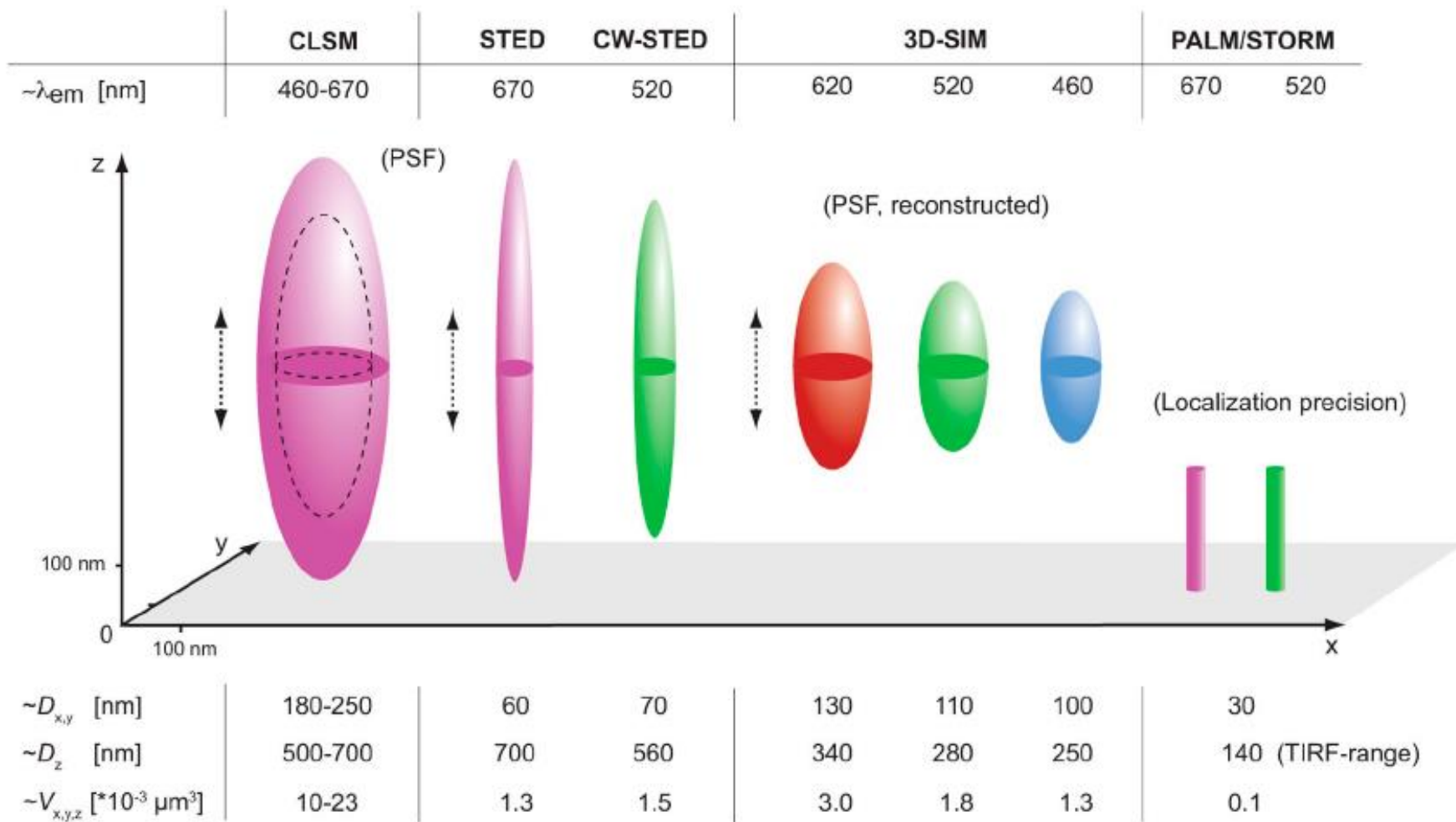
SIM

STED

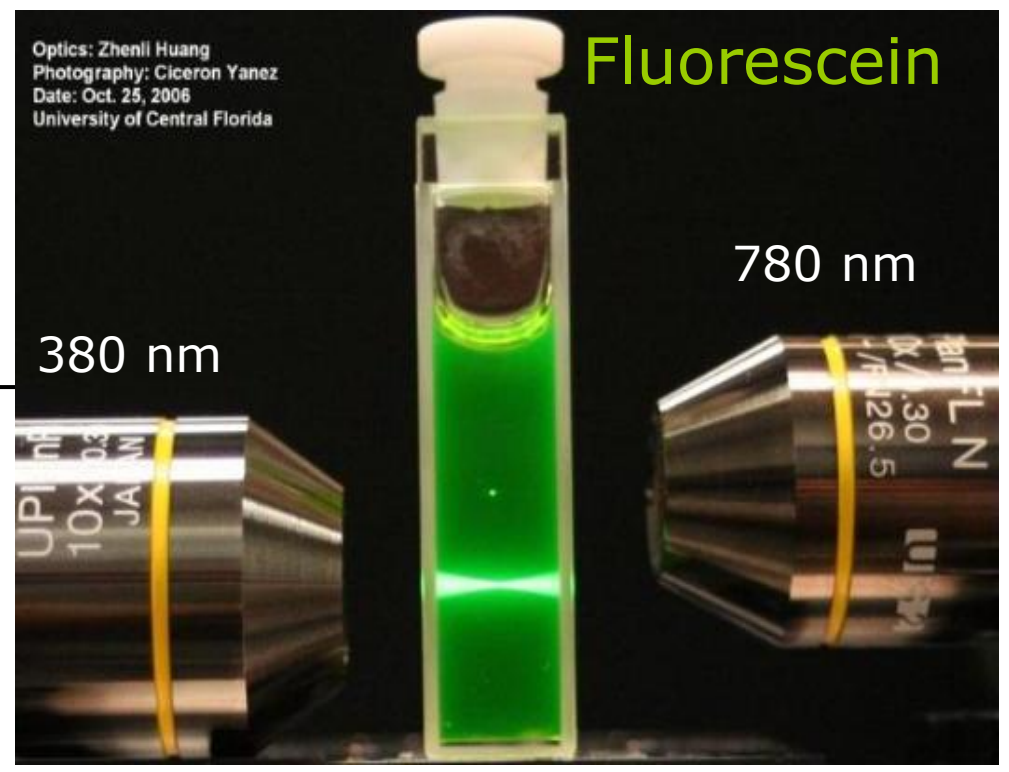
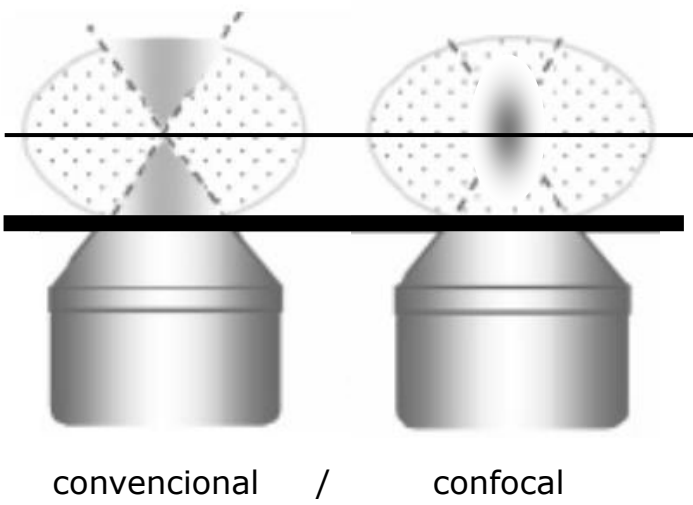
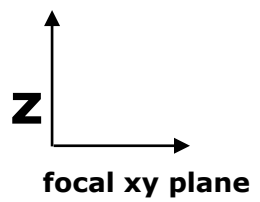
4- $\pi$

PALM

# |-> PSF overview



| Best localization: confocal microscopy



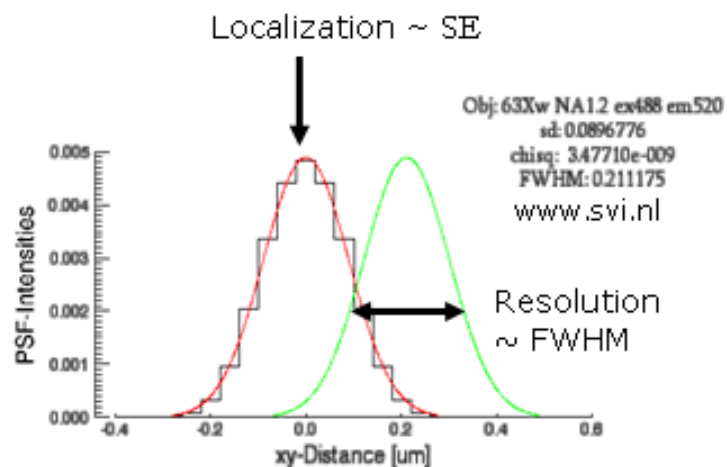
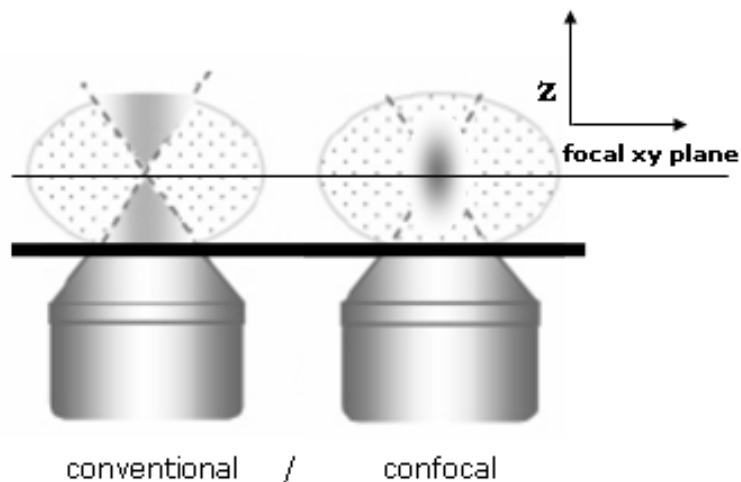
## | Diffraction limited microscopy

E. Abbe († 1905)

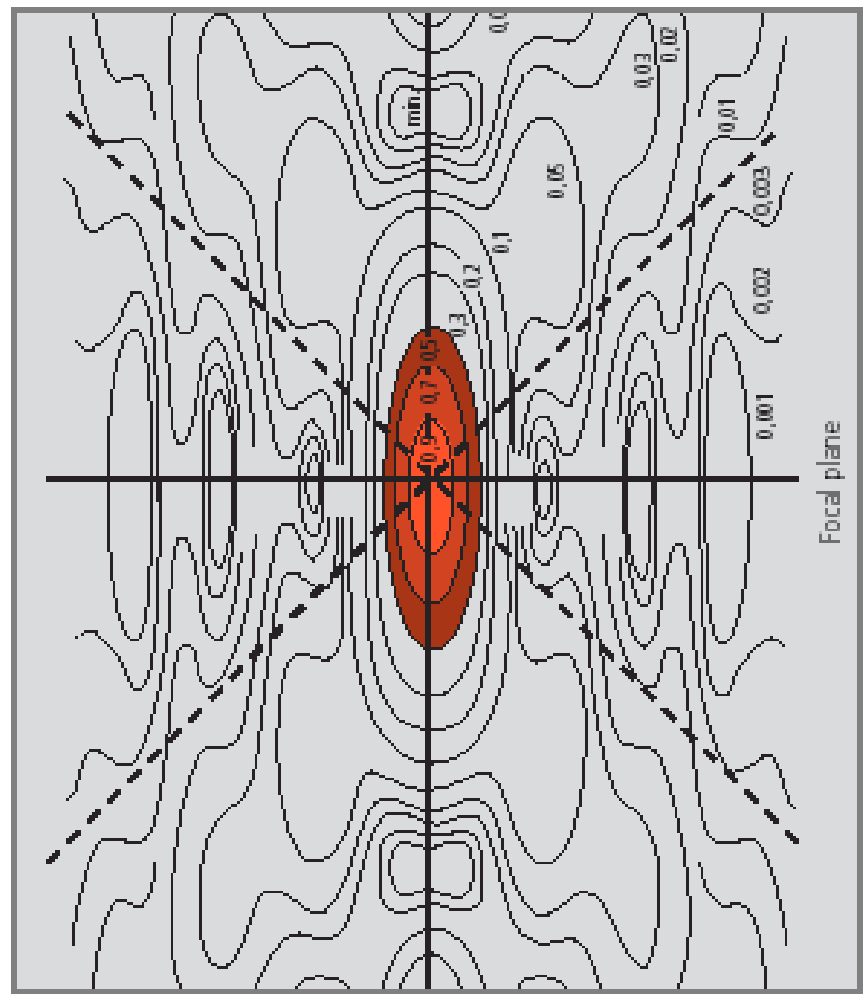
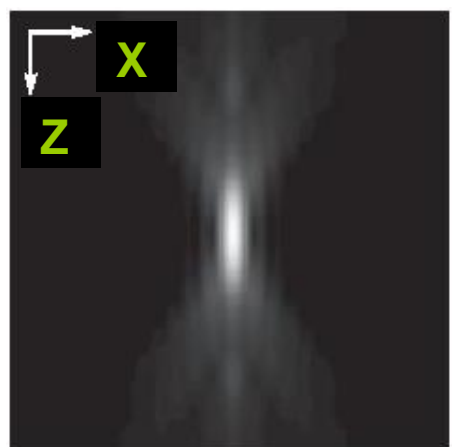
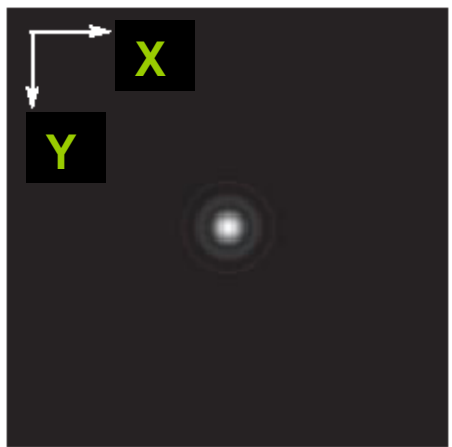


$\lambda / 2 \cdot NA \sim \lambda / 2$  Resolution (Full Width at Half Maximum, FWHM)

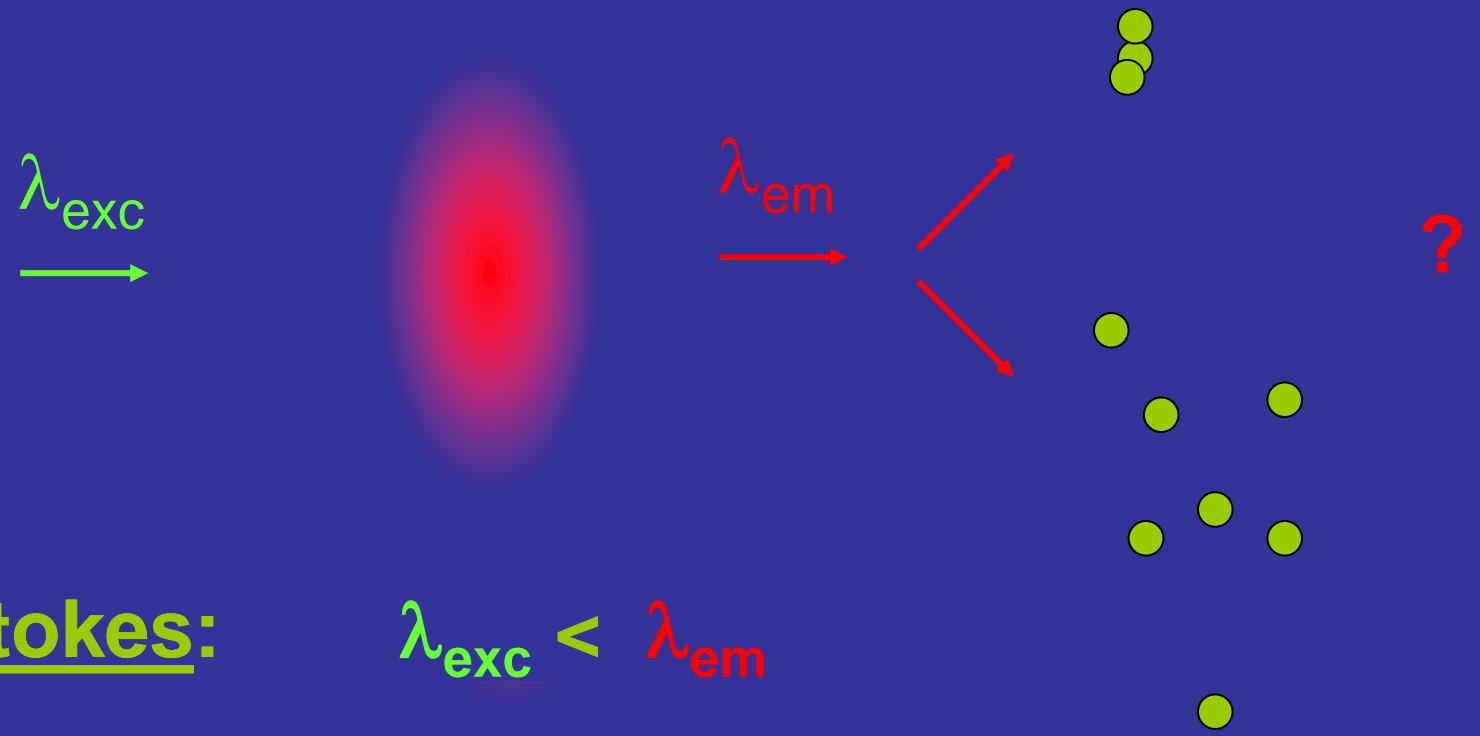
FWHM /  $N^{1/2}$  Localization, N number of photons



# | -> PSF



# | -> Convolution





# |-> Convolution

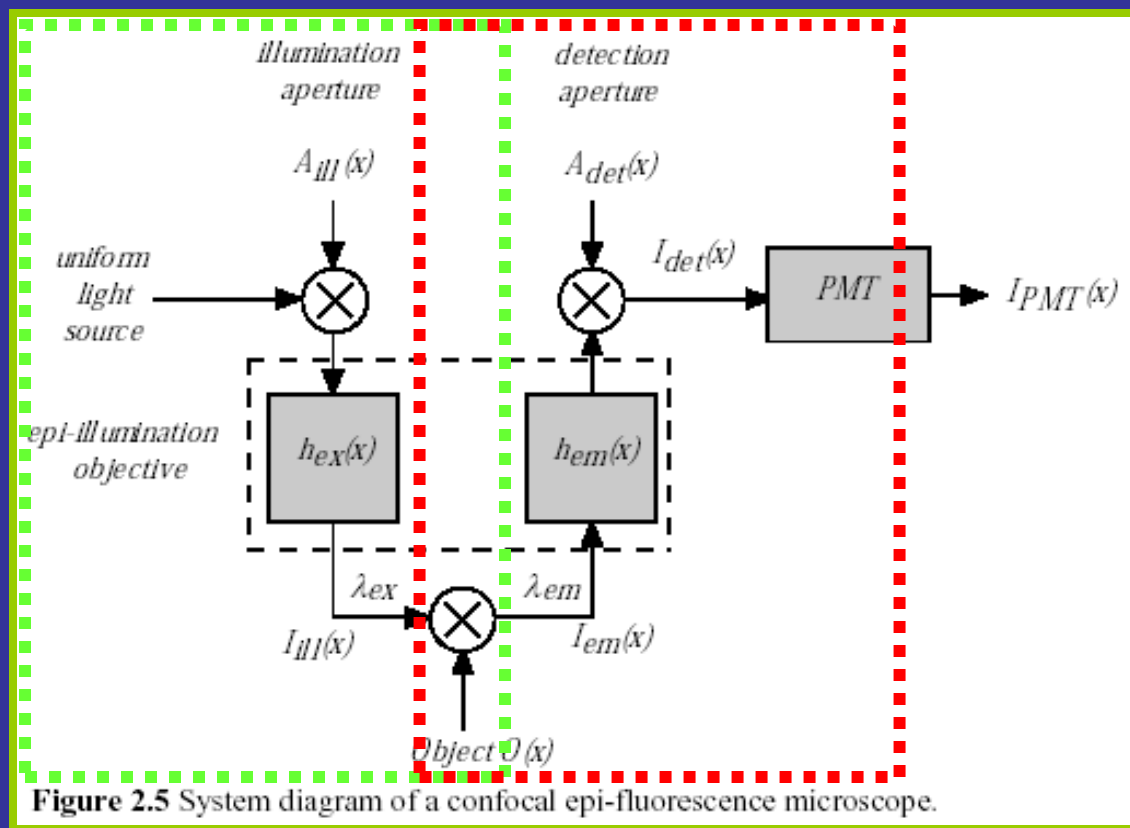
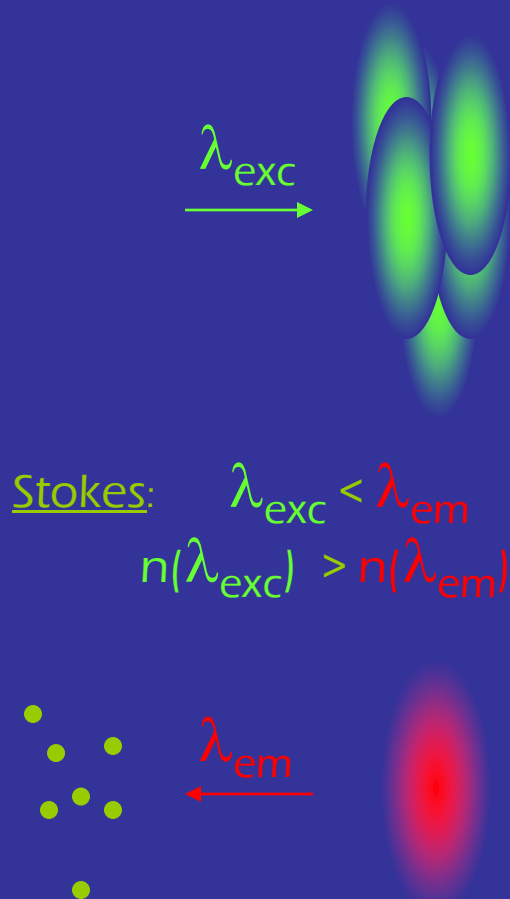
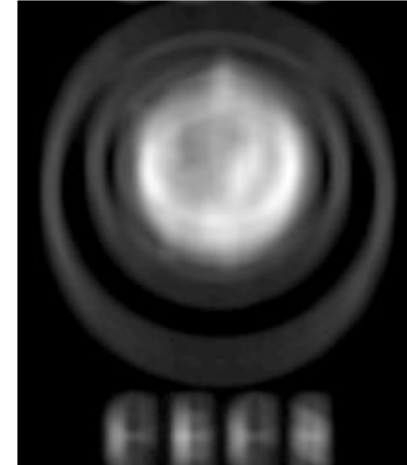
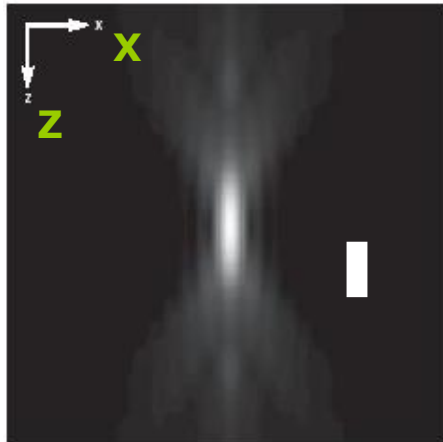


Figure 2.5 System diagram of a confocal epi-fluorescence microscope.

# | -> Convolution



**PSF:**  
 $\Delta xy \sim 500 \text{ nm}$  |  $\Delta z \sim 1500 \text{ nm}$

*PSF: Point Spread Function*

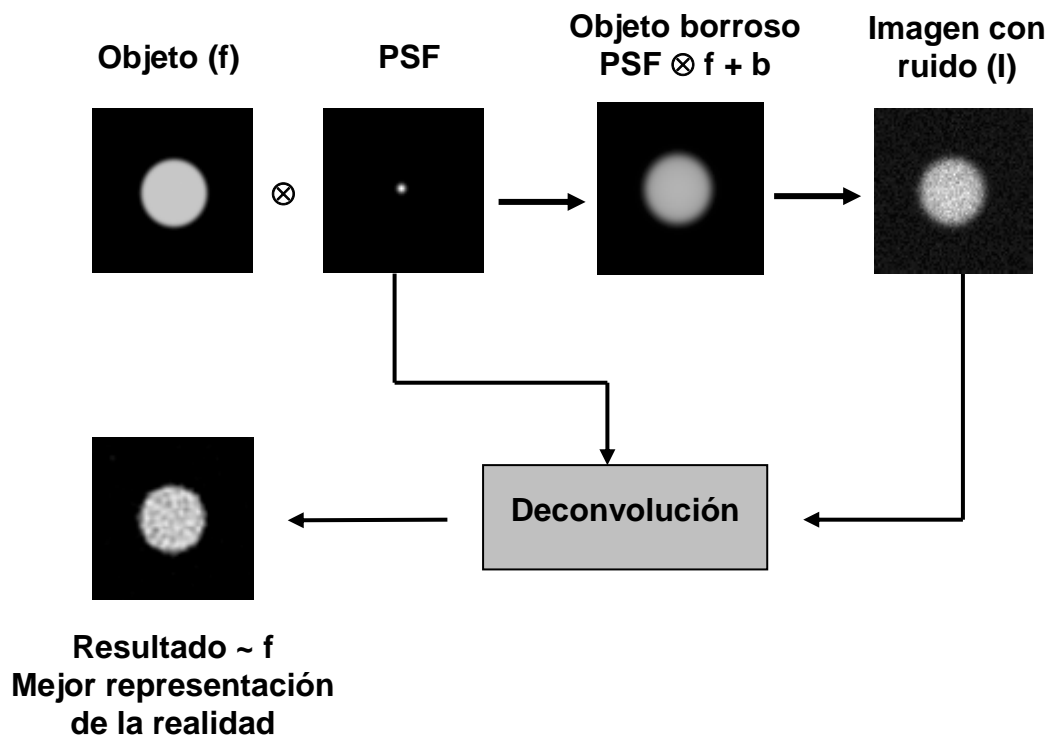
*f: Object Function*

*b: Offset Function*

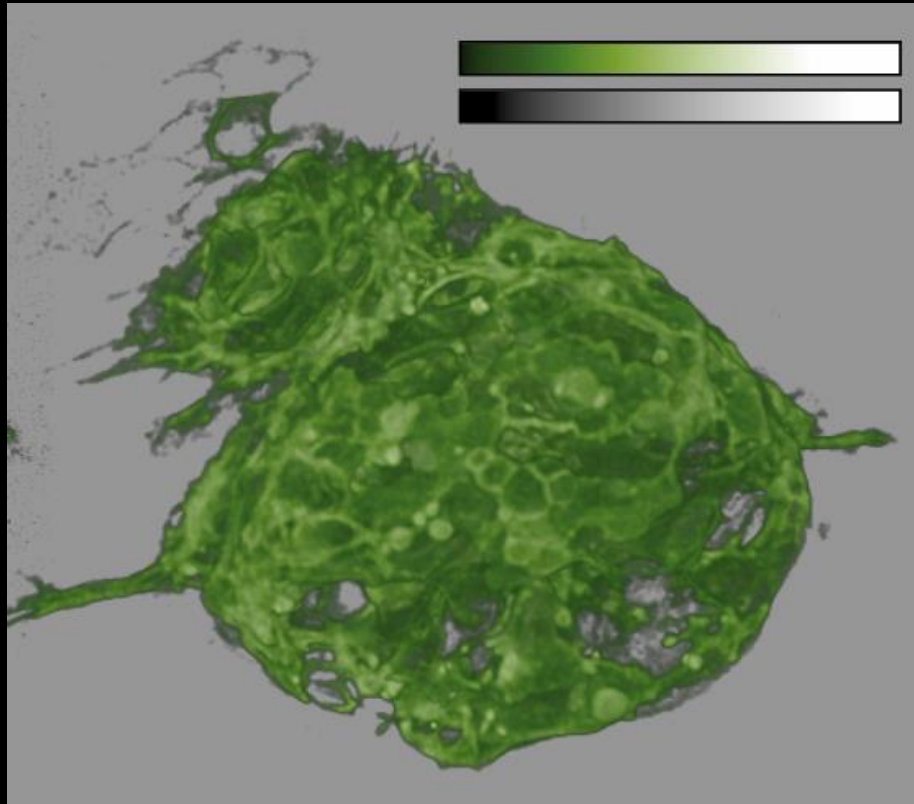
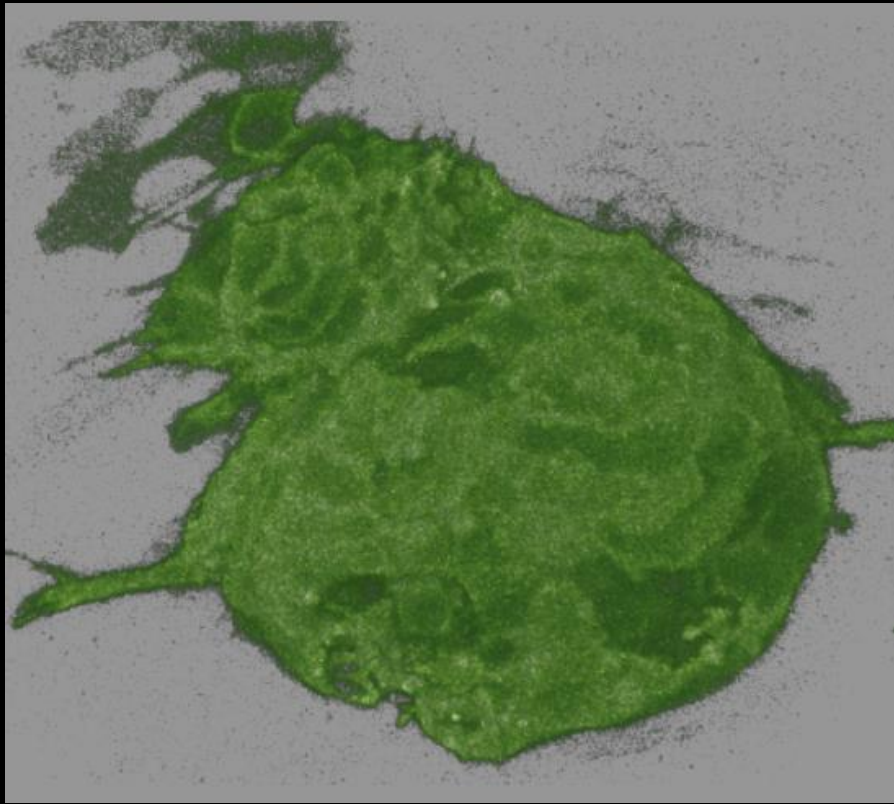
*I: Image Matrix*

*N: Noise Function*

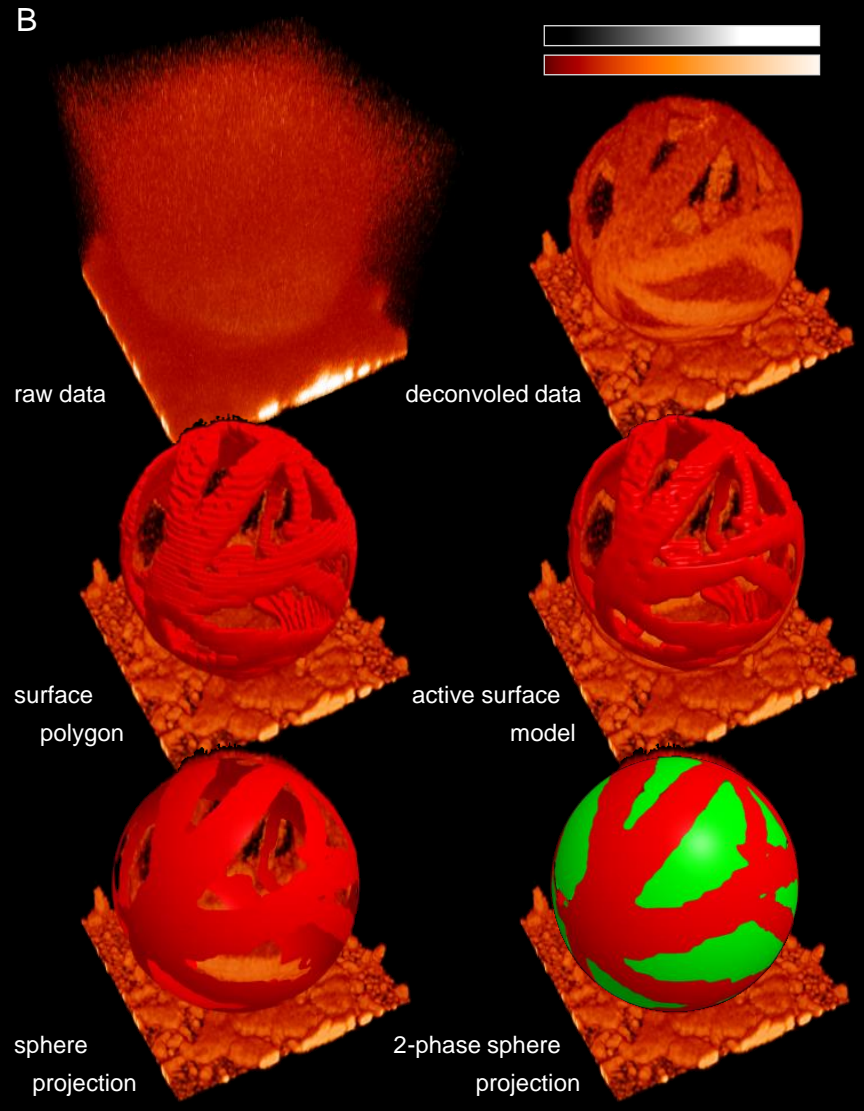
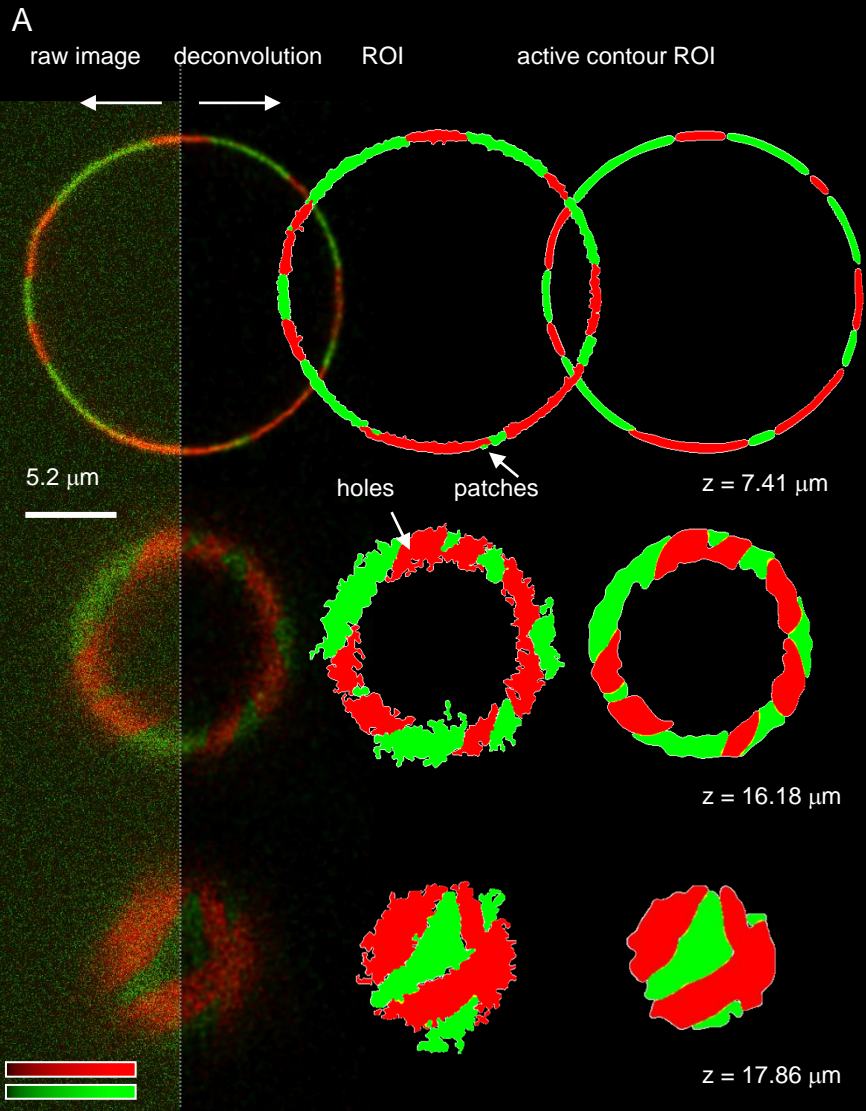
$$N(PSF(x, y, z) \otimes f(x, y, z) + b(x, y, z)) = I(x, y, z)$$



# |-> Deconvolution



# | -> Deconvolution



*PSF: Point Spread Function*

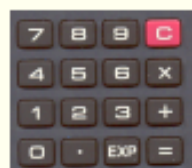
*f: Object Function*

*b: Offset Function*

*I: Image Matrix*

*N: Noise Function*

$$N / (PSF(x, y, z) \otimes f(x, y, z) + b(x, y, z)) = I(x, y, z)$$



## Calculator

[Numerical aperture](#)

[Excitation wavelength](#)

(nm)

[Emission wavelength](#)

(nm)

[Number of excitation photons](#)

[Backprojected pinhole radius](#)

(nm)

[B.P. distance between pinholes](#)

Only for Nipkow disks (μm)

[Lens medium refractive index](#)

[Specimen medium refractive index](#)

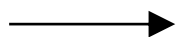
[Acquisition depth](#)

(μm)

Calculate also PSF

- confocal
- widefield
- nipkow
- 4Pi

Select one





*PSF: Point Spread Function*

*f: Object Function*

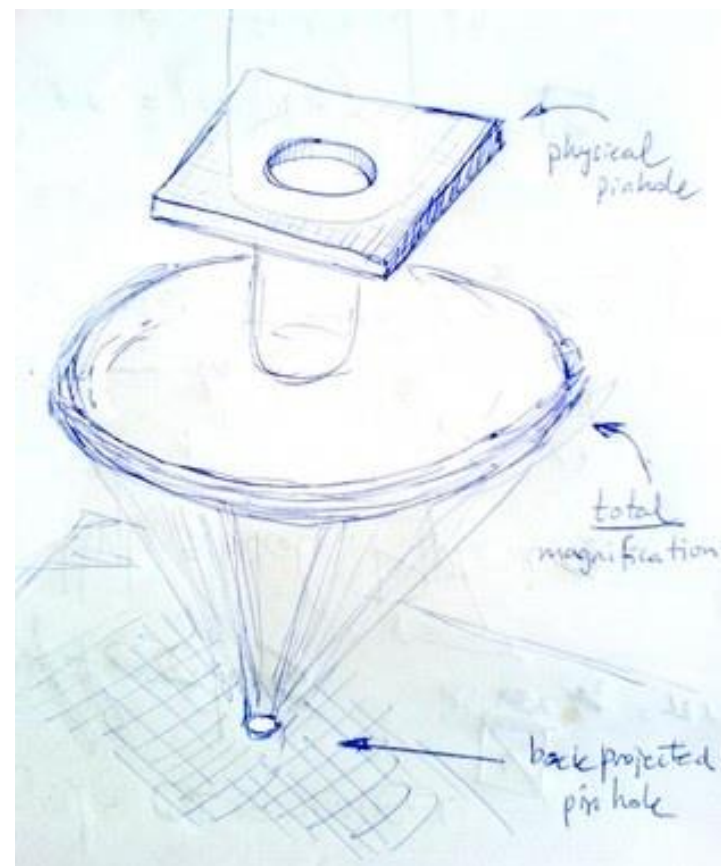
*b: Offset Function*

*I: Image Matrix*

*N: Noise Function*

$$N / (PSF(x, y, z) \otimes f(x, y, z) + b(x, y, z)) = I(x, y, z)$$

## Backprojected confocal pinhole



<http://support.svi.nl/wiki/NyquistCalculator>

*PSF: Point Spread Function*

*f: Object Function*

*b: Offset Function*

*I: Image Matrix*

*N: Noise Function*

$$N(PSF(x, y, z) \otimes f(x, y, z) + b(x, y, z)) = I(x, y, z)$$

## Biorad

- [Biorad MRC 500, 600 and 1024](#)
- [Biorad Radiance](#)

## Leica

- [Leica confocals TCS 4d, SP1 and NT](#)
- [Leica confocal SP2](#)
- [Leica confocal SP5](#)

## Nikon

- [TE2000-E with the C1 scanning head](#)

## Olympus

- [Olympus FV300](#)
- [Olympus FV500](#)
- [Olympus FV1000](#)

## Zeiss

- [Zeiss LSM410 inverted](#)
- [Zeiss LSM510](#)



# | -> Noise



Literature: eg. Noise Theory and Application to Physics: Philippe Réfrégier, Springer

*PSF: Point Spread Function*

*f: Object Function*

*b: Offset Function*

*I: Image Matrix*

*N: Noise Function*

$$N(PSF(x, y, z) \otimes f(x, y, z) + b(x, y, z)) = I(x, y, z)$$

$$P(p, \mu) = \frac{\mu^p}{p!} \cdot e^{-\mu}$$

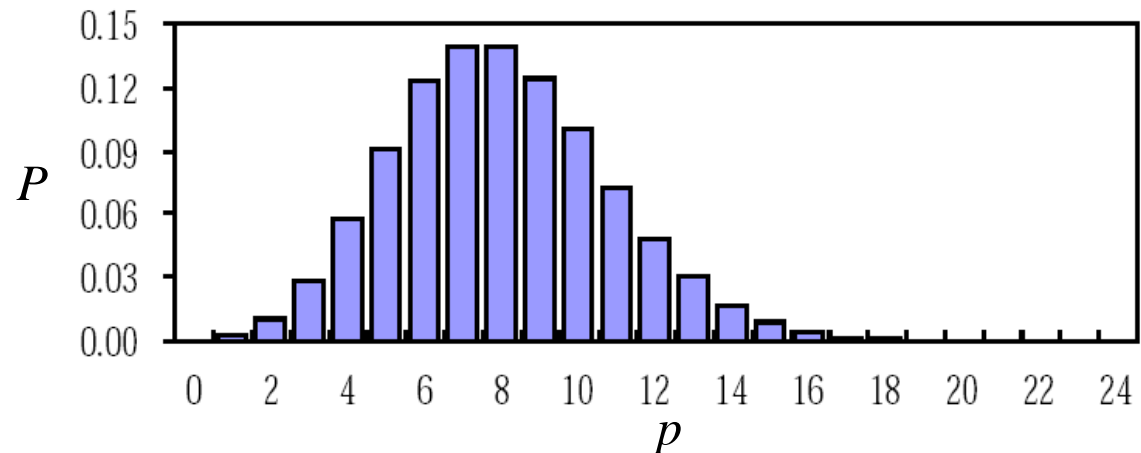
$$1. \bar{p} = \mu = \sigma^2, sd = \sigma = \sqrt{\bar{p}} = \sqrt{\mu}$$

$$2. \text{counting} : \bar{p} \pm \sqrt{\bar{p}}$$

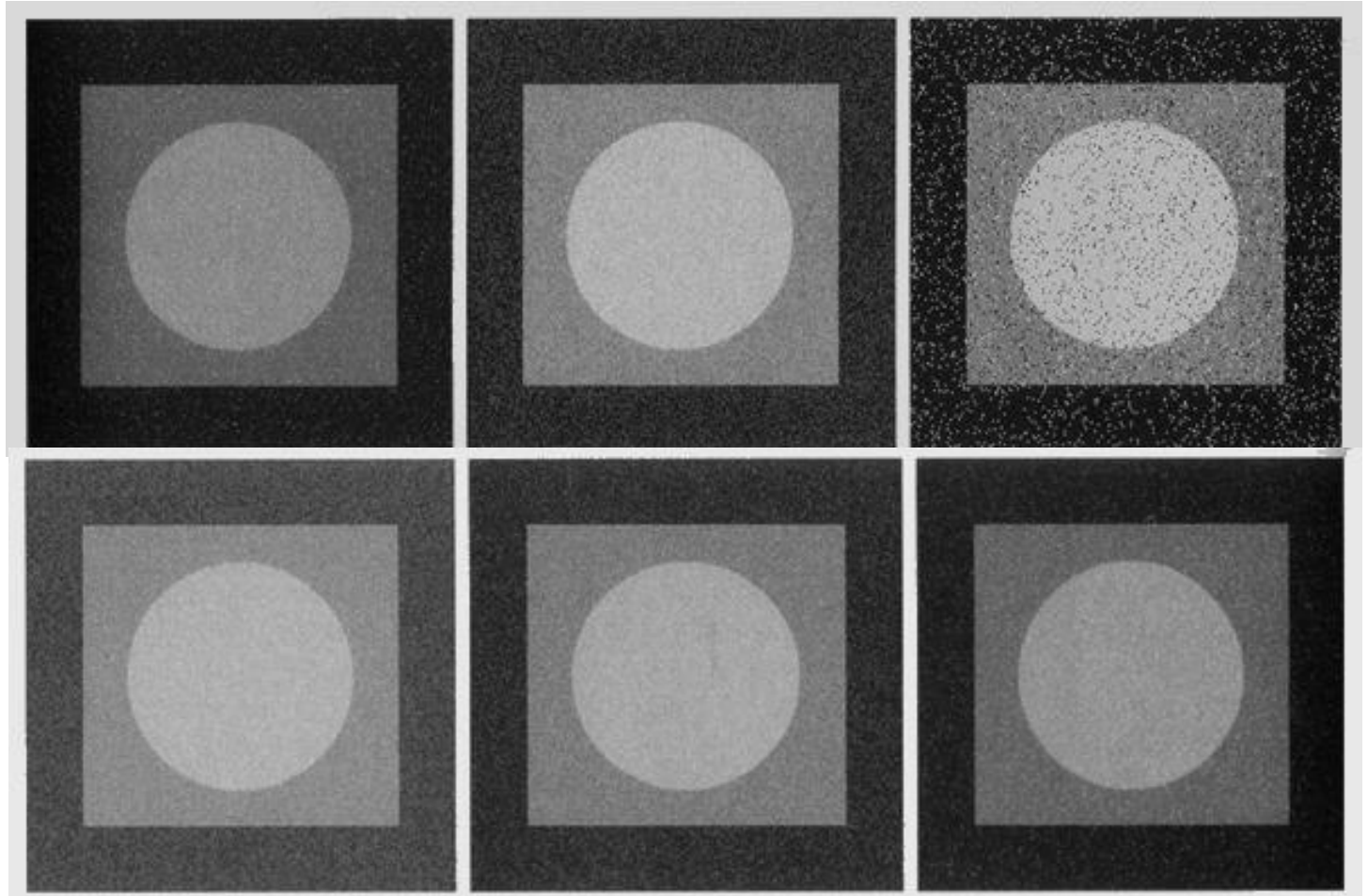
$$3. \text{Poisson}(\text{discrete}) \rightarrow \text{Gauss}(\text{continuous}) : \mu \rightarrow \infty$$

- *Black Body Irradiation (Poisson)*

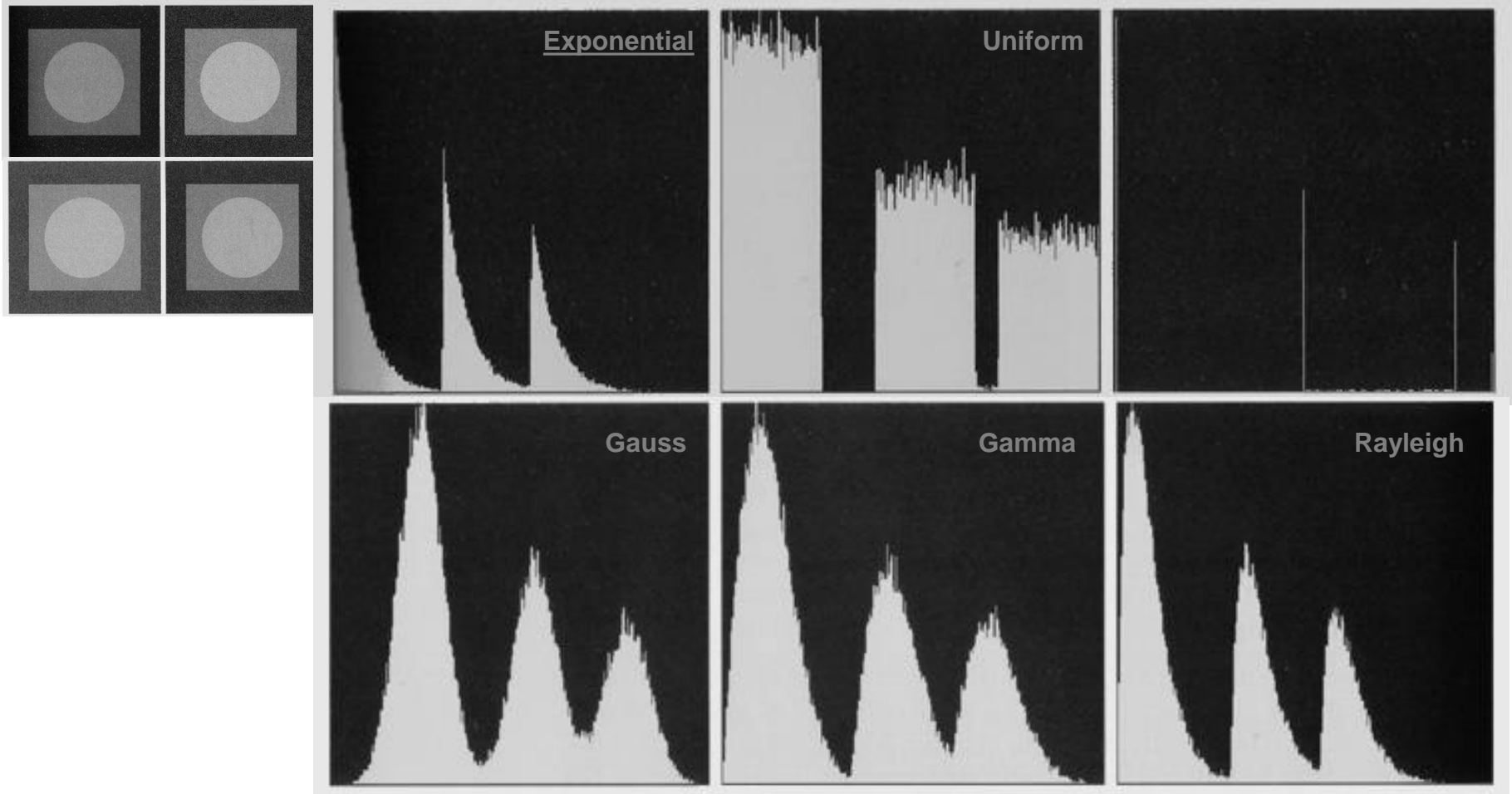
- *Detector Noise (Gauss)*



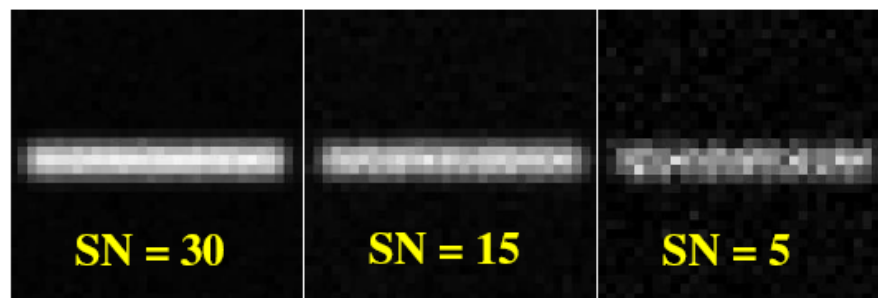
# | -> Noise



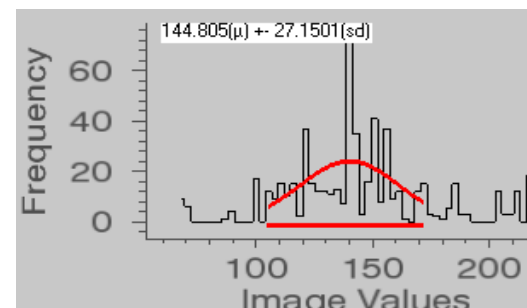
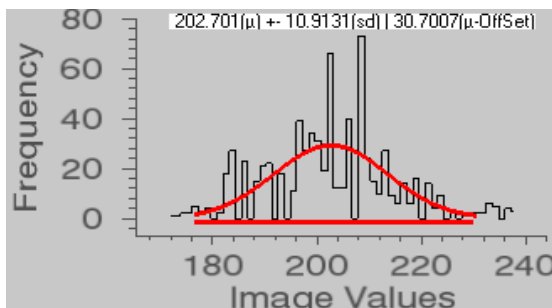
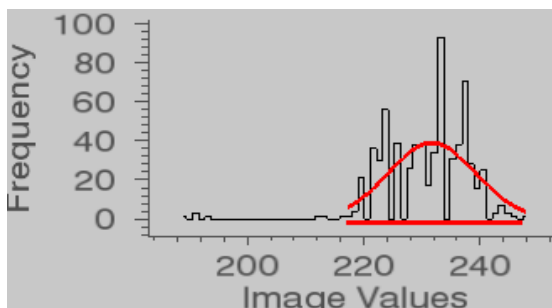
# | -> Noise



The Signal to Noise ratio (SN) is a number not always easy to estimate. The easiest way to obtain some figures is to look at the textures of bright areas in your object image. In the figure at left you see examples of such textures obtained from originally the same object image to which various levels of poisson noise were added.



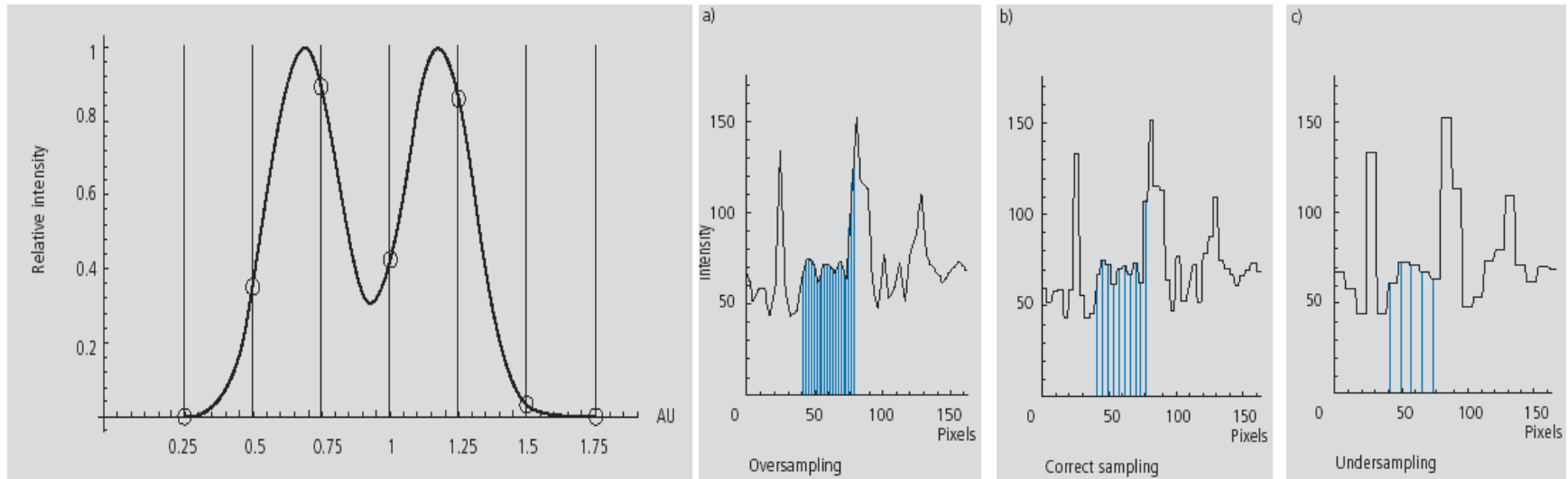
**Figure 9. Images with different generated noise levels**



$$SNR = \frac{\bar{I}}{\sigma} = \frac{\bar{I}}{\sqrt{\sigma^2}} = \frac{229}{7.5}$$

$$SNR = \frac{\bar{I}}{\sigma} = \frac{\bar{I}}{\sqrt{\sigma^2}} = \frac{200}{10}$$

$$SNR = \frac{\bar{I}}{\sigma} = \frac{\bar{I}}{\sqrt{\sigma^2}} = \frac{139}{27}$$



- Undersampling loses structures.
- Oversampling waists memory/computation time.

The 'Nyquist /Shannon Theorem' or 'Sampling Theorem' for the digital sampling of analogue signals suggests a Nyquist rate  $NR \geq 2v$  ?

**! Diffraction theory calculates lateral  $NR \sim 20 \text{ pixel}/\mu\text{m} (\sim 50 \text{ nm}/\text{pixel})$  !**  
**... axial  $NR \sim (\sim 150 \text{ nm}/\text{pixel})$**



*PSF: Point Spread Function*

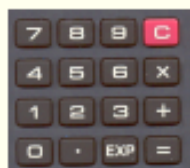
*f: Object Function*

*b: Offset Function*

*I: Image Matrix*

*N: Noise Function*

$$N(PSF(x, y, z) \otimes f(x, y, z) + b(x, y, z)) = I(x, y, z)$$



## Calculator

[Numerical aperture](#)

[Excitation wavelength](#)

 (nm)

[Emission wavelength](#)

 (nm)

[Number of excitation photons](#)

[Backprojected pinhole radius](#)

 (nm)

[B.P. distance between pinholes](#)

 Only for Nipkow disks (μm)

[Lens medium refractive index](#)

[Specimen medium refractive index](#)

[Acquisition depth](#)

 (μm)

Calculate also PSF

- confocal
- widefield
- nipkow
- 4Pi

Select one

*PSF: Point Spread Function*

*f: Object Function*

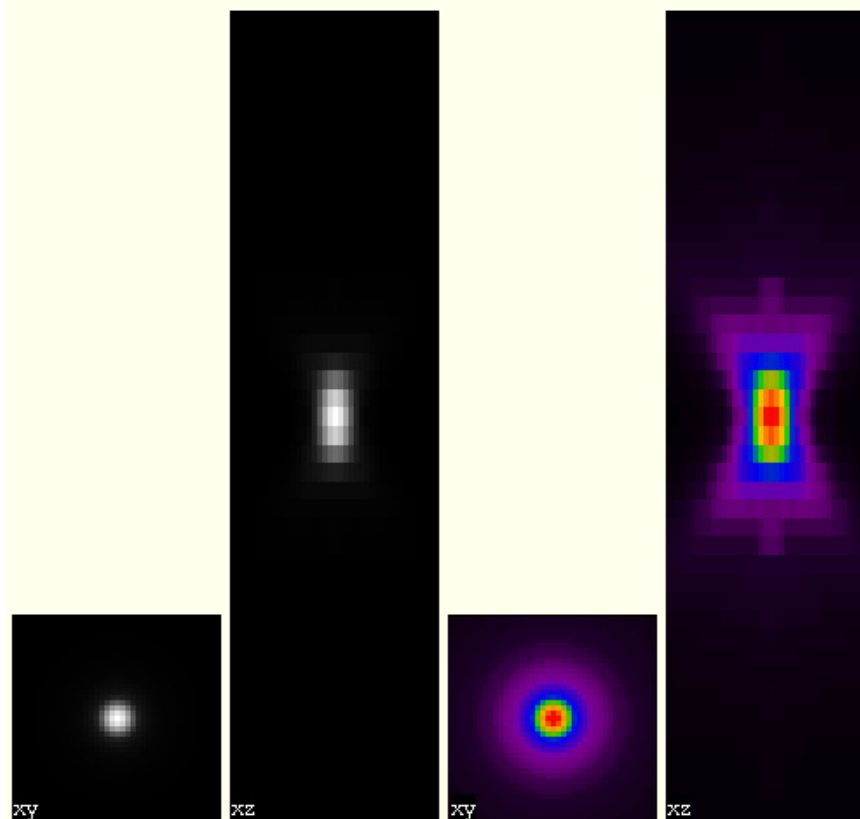
*b: Offset Function*

*I: Image Matrix*

*N: Noise Function*

$$N(PSF(x, y, z) \otimes f(x, y, z) + b(x, y, z)) = I(x, y, z)$$

Nyquist sampling (x,y,z in nm): 46, 46, 165





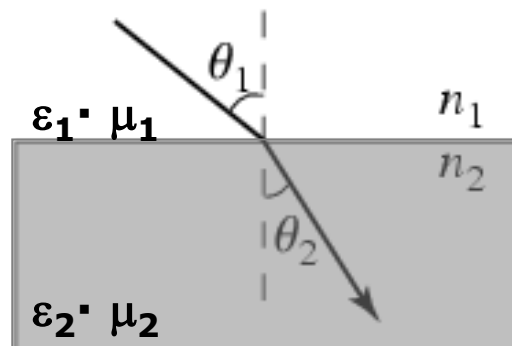
**Index of refraction:  $n = (\varepsilon \cdot \mu)^{1/2} = c/v$ ,**

*$\varepsilon$  electric permittivity and  $\mu$  magnetic permeability.*

## Snell's Law:

$$\sin \theta_1 n_1 = \sin \theta_2 n_2$$

- 1.518 [Zeiss Oil]
- 1.33 [Water]
- 1.0008 [Air]



## Refractive Index:

$$RI = n_1/n_2 = v_2/v_1$$

## Snell's Law:

$$\sin \theta_1 n_1 = \sin \theta_2 n_2$$

$$n = n(\lambda) !$$

- **1.518 [Zeiss]**
- **1.33 [Water]**
- **1.0008 [Air]**

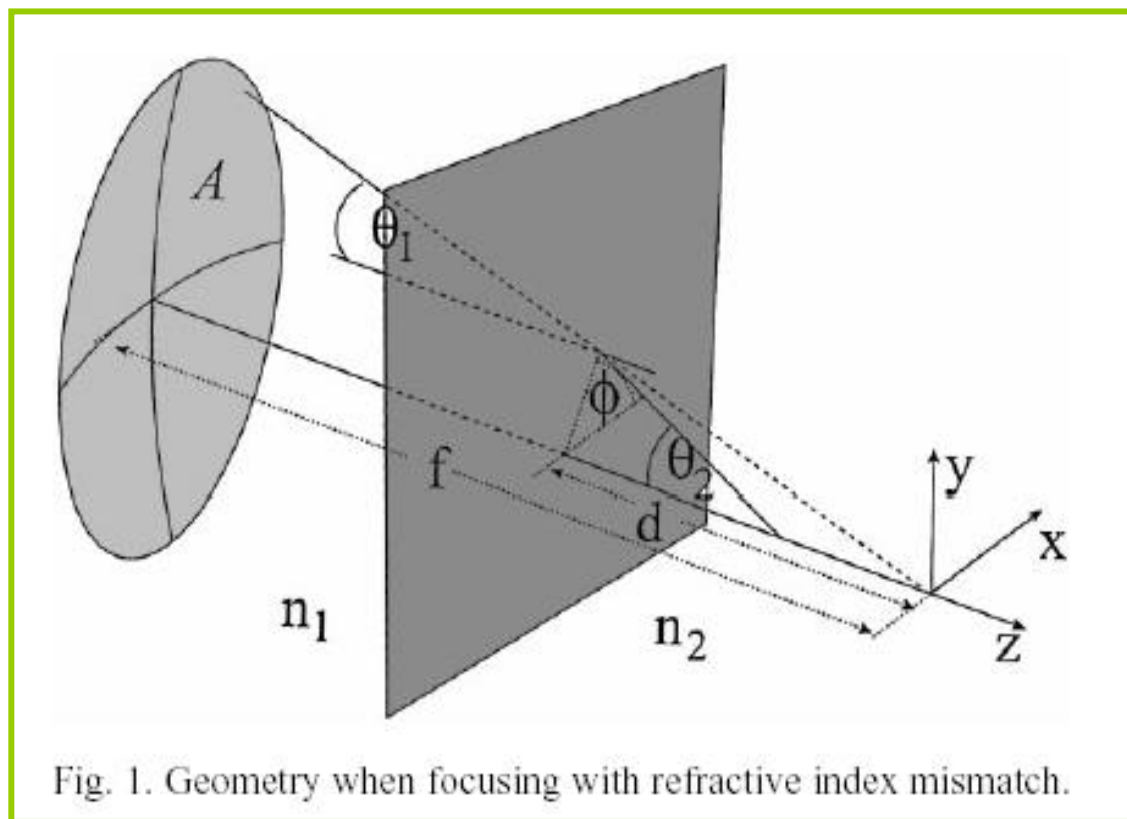
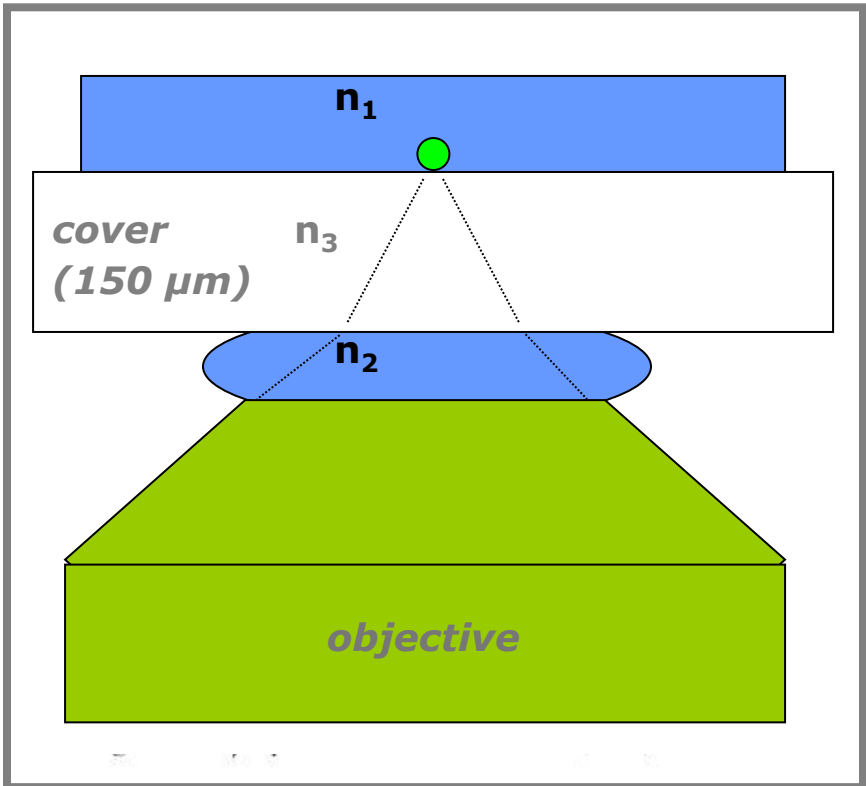


Fig. 1. Geometry when focusing with refractive index mismatch.

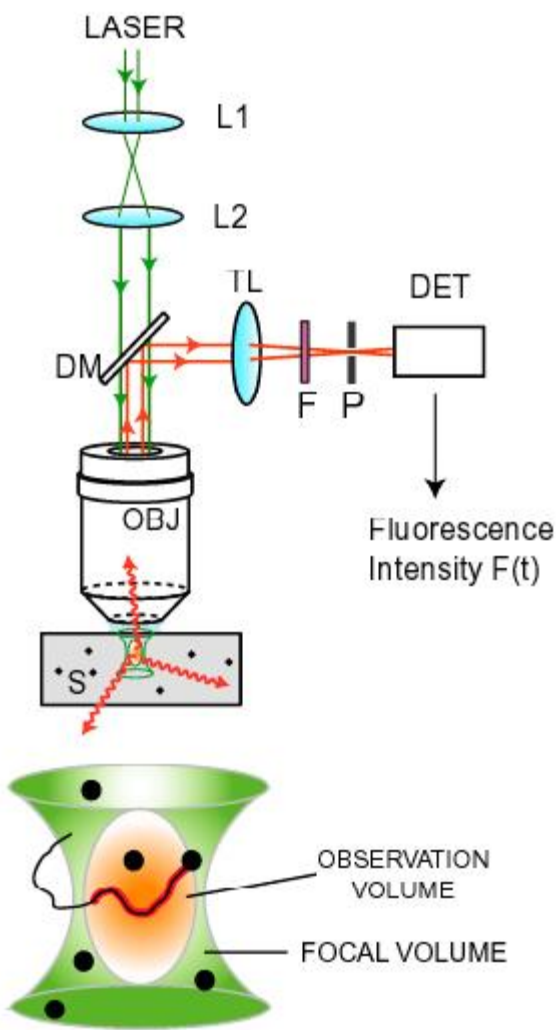
*(Egner et al 1998)*

● **Micro-esfera:  $\varnothing = 6 \mu\text{m}$**



*agua/aceite* -- *aceite/aceite*  
 $n_1 \neq n_2$        $n_1 = n_2$

**Ley de Snell:  $n_i \cdot \sin\theta_i = n_k \cdot \sin\theta_k$**   
 $n = n(\lambda) !$



The observation volume (femtoliter) defined by the Point Spread Function must be considered as a mini-spectrofluorimeter.

1. You need to consider the Offset  $I(0)$  in order to calibrate your signal  $I(0) \geq 0$  !
2. Never saturate the signal:  $I \leq I_{\max}$  (255 for 8 bit) !

